CHEMICAL PROFILING OF SIX SAMPLES OF BRAZILIAN PROPOLIS

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INTRODUCTION

The word propolis derives from the Greek pro (in defense of, or in front of) and polis (city), implying a product useful for the defense of the hive. It is a complex mixture of compounds with resinous aspect, made by Apis mellifera bees from plant resins and beeswax. Its chemical composition depends on the plant or plants from which the resin is collected and, consequently, on the geographical location of the hive.1-3 Propolis samples have been shown to exhibit many biological activities, such as antimicrobial,4 antiviral,5 antiinflammatory,6 antiprotozoan,7 antitumoral,8,9 and antioxidant actions.10,11

Many types of propolis, comprising a wide diversity of botanical sources, have been described. The main sources of European propolis are buds of poplars (Populus spp.).2 In Cuba and Venezuela bees collect resins from Clusia spp.3 In Brazil, there are at least four distinct resin sources for propolis production: Baccharis dracunculifolia, the alecrim-do-campo plant (Brazilian green propolis),3,13 Dalbergia ecastophyllum (Brazilian red propolis),14 Hyptis divaricata (Brazilian brown propolis) and Populus alba (poplar type propolis).15 Recently, new propolis types and botanical sources have been described, such as Pacific propolis (plant source: Macaranga spp.),18 Mediterranean propolis (plant source: conifers)17 and Brazilian Amazonian propolis (plant source: probably Clusia sp.).8,19

Among Brazilian propolis, the most exported and intensively studied is the green type.2 It is composed mainly of prenylated phenylpropanoids, such as artepillin C (3,5-diprenyl-4-hydroxyxcinamic acid)3,20 and 3-prenylcinnamic acid allyl ester, both compounds assumed as markers of green propolis.29 Flavonoids, such as kaempferide, are present, although not as major constituents.1,2,21 Terpenoids and benzoic acids may also be found in green propolis.3,22 Propolis from the south of Brazil has been regarded as derived either from poplars14 or Araucaria.23 Amounts of propolis constituents have rarely been investigated using standardization, either internal or external. The determination of major propolis constituents may be crucial for standardization and chemical quality control.24

The aim of the present study was to compare the chemical composition of four samples of Brazilian green propolis from Minas Gerais (southeast) and two from Paraná (south of Brazil). The former state is the geographical center of distribution of green propolis, while Paraná state lies on the south border of this zone of distribution. This study also sought to determine the contents of relevant constituents of the six samples, using external standardization.

EXPERIMENTAL

Propolis samples and extraction

Samples A and B were produced in the municipality of Esmeraldas, state of Minas Gerais. Samples C and D came from the municipality of Três Pontas, state of Minas Gerais. Samples E and F came from the municipality of União da Vitória, state of Paraná. Successive extractions were carried out in Soxhlet with 5 g of each sample and the solvents hexane, chloroform (CHCl₃), ethyl acetate (EtOAc) and methanol (MeOH). The extracts were concentrated to dryness under reduced pressure.

Derivatization

A 30 μL volume of a 10 mg/mL CHCl₃ solution of the CHCl₃ extracts was treated with 4 mL of a 5% MeOH solution of H₂SO₄ and 2 mL of toluene. The mixture was left standing in a steam bath at 80 °C for 4 h. Extraction was then performed with 2 mL of 0.5 M NaCl solution and 1 mL of methylene chloride. The mixture was vigorously stirred and then centrifuged at 5000 rpm for 5 min. The organic phase was collected and the residue extracted with methylene chloride twice using the same procedure. The pooled organic phases were washed 3 times with 0.5 M NaCl and the aqueous phase was discarded. The extract containing the derivatized products was treated with anhydrous Na₂SO₄ and concentrated under a N₂ flow.

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**Chemical composition**

Hexane and derivatized CHCl₃ extracts were analyzed by GC/MS according to Negri et al. Identification of the compounds was accomplished using computer-based searches of commercial libraries and literature data.

The EtOAc and MeOH extracts were dissolved in MeOH at the concentration of 10 mg/mL. Both extracts from all 6 samples were analyzed by injecting 10 µL of the MeOH solutions into an HPLC chromatograph equipped with a C18 RP Luna Phenomenex column (4.6 x 250 mm, 5 µm). The mobile phase used contained 0.1% acetic acid and MeOH, with a constant 0.5 mL/min flow. The HPLC-DAD-ESI-MS system was a DADSPD-M10A VP Shimadzu equipped with a photodiode array detector, coupled to an Esquire 3000 Plus, Bruker Daltonics. The mass detector was a quadrupole ion trap equipped with an atmospheric pressure ionization source through electrospray ionization interface. The mobile phase flow was 0.5 mL/min and the gradient used comprised MeOH 20 to 40%, from 0 to 10 min; MeOH 40 to 60%, from 10 to 20 min; MeOH 60 to 80%, from 20 to 30 min; MeOH 80 to 95%, from 30 to 37 min; and MeOH 95%, from 37 to 45 min. Detection was accomplished at 270 and 300 nm. Mass spectra were obtained using a negative ESI source voltage of ~40 V and a capillary offset voltage of 4500 V. Nebulization was aided with coaxial nitrogen sheath gas, provided at 27 psi pressure. Temperature of the dry gas was 130 °C and the flow was 4 L/min. A counter current nitrogen flow was set at 7 L/min and capillary temperature at 320 °C, to assist desolvation. Mass spectra were recorded over the range 50-700 m/z. The identification of sample constituents was based on their UV absorbance band and on cross-comparison of mass spectra data with literature data.

**Quantification of constituents**

The contents of the main compounds were determined by HPLC analysis and external standardization, using 10 µL of MeOH solutions of the EtOAc and MeOH extracts. The solutions were injected into an HP 1090 HPLC apparatus, equipped with a reverse phase C18 column (4.6 x 250 mm, 5 µm), using the gradient described in the previous section. The amount of compounds was estimated on the basis of the areas under the corresponding peaks and standard curves prepared with quercetin (for flavonoids), p-coumaric acid (phenylpropanoids) and chlorogenic acid (caffeoylquinic acids). The contents of the compounds were expressed as mg per g of crude propolis.

**RESULTS AND DISCUSSION**

The compounds identified by GC/MS are listed in Table 1. Although with different peak intensities, all compounds were detected in the 6 samples analyzed. Compounds 3 and 6 (palmitic and stearic acids, respectively) are common constituents of natural waxes. Compound 2 is a simple phenol which has been reported from Brazilian green propolis, sometimes as one of the major constituents. With the exception of 1 (benzenepropanoic acid), all other compounds were phenylpropanoids. Compounds 4 (an allyl ester), 5 (drupanin), as well as 10, contain one prenyl group. The same holds for the chromanes 7 and 8, both bearing a prenyl group involved in

**Table 1. Constituents of six samples of Brazilian propolis characterized by GC-ESI-MS. A-D: samples from the state of Minas Gerais (southeast Brazil); E and F: samples from the state of Paraná (south Brazil)**

<table>
<thead>
<tr>
<th>Peak</th>
<th>RT</th>
<th>Molecular ion and fragments</th>
<th>Proposed compounds</th>
<th>Relative amount (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>6.78</td>
<td>150 (40, C₇H₈O₇), 104 (70), 91(100), 77(30)</td>
<td>benzenepropanoic acid[12,23]*</td>
<td>26.94 23.77 18.5 18.2 15.52 26.94</td>
</tr>
<tr>
<td>2</td>
<td>10.35</td>
<td>188 (70, C₇H₈O₇), 133 (100), 104 (30), 91 (30), 77 (30)</td>
<td>p-vinyl-α-prenylphenol[12,23]*</td>
<td>14.77</td>
</tr>
<tr>
<td>3</td>
<td>13.42</td>
<td>270 (1, C₁₇H₃₃O₂), 143 (30), 87(70), 74 (100), 41 (98)</td>
<td>palmitic acid methyl ester[22,25]*</td>
<td>0.1 26.90 0.1 3.22 0.1 0.1</td>
</tr>
<tr>
<td>4</td>
<td>15.05</td>
<td>256 (C₆H₁₂O₈), 185 (70), 145 (100), 91 (40), 77 (20), 69 (30)</td>
<td>3-prenylcinnamic acid allyl ester[23,25]*</td>
<td>23.99 20.95 32.42 22.58 4.09 9.73</td>
</tr>
<tr>
<td>5</td>
<td>15.46</td>
<td>246 (70, C₁₇H₃₃O₂), 191 (100), 171 (20), 131 (23)</td>
<td>4-hydroxy-3-prenylcinnamic acid (drupanin) methyl ester[22,25,26,27,28]*</td>
<td>0.1 26.94 29.59 24.54 14.84 17.57</td>
</tr>
<tr>
<td>6</td>
<td>16.31</td>
<td>298 (4, C₁₇H₃₃O₂), 143 (30), 87(70), 74(100)</td>
<td>stearic acid methyl ester[22,25]*</td>
<td>0.1 0.1 &lt; 0.1 2.67 1.56 0.1</td>
</tr>
<tr>
<td>7</td>
<td>17.48</td>
<td>312 (14, C₁₇H₃₃O₂), 297 (100)</td>
<td>2,2-dimethyl-8-prenylchromene-6-propenoic acid methyl ester[22,25,26,27,28]*</td>
<td>3.38 0.1 5.13 4.96 1.53 0.1</td>
</tr>
<tr>
<td>8</td>
<td>19.11</td>
<td>330 (100, C₁₇H₃₃O₂), 297 (30), 272 (50), 225 (60), 197 (50), 171 (50).</td>
<td>3-hydroxy-2,2-dimethyl-8-prenylchromene-6-propenoic acid methyl ester[22,25,26,27,28]*</td>
<td>2.23 0.1 30.37 6.39 1.97 3.76</td>
</tr>
<tr>
<td>9</td>
<td>19.23</td>
<td>314 (68, C₁₇H₃₃O₂), 259 (100), 245 (54), 211 (38), 203 (90).</td>
<td>4-hydroxy-3,5-diprenylcinnamic acid (artepillin C) methyl ester[22,25,26,27,28]*</td>
<td>3.65 4.54 5.13 2.53 1.91 3.47</td>
</tr>
<tr>
<td>10</td>
<td>21.43</td>
<td>330 (100, C₁₇H₃₃O₂), 297 (50), 259 (70), 228 (30), 203 (60)</td>
<td>3-prenyl-4-(2-methylpropionyl-ox)-cinnamic acid[22,23]</td>
<td>0.1 0.1 1.91 0.1 2.66</td>
</tr>
</tbody>
</table>

* = compounds detected in non-derivatized hexane extracts; *= compounds detected in derivatized chloroform extracts
Table 2. Constituents of six samples of Brazilian propolis characterized and quantified by HPLC-ESI-MS. A-D: samples from the state of Minas Gerais (southeast Brazil); E and F: samples from the state of Paraná (south Brazil).

<table>
<thead>
<tr>
<th>Peak</th>
<th>RT (min)</th>
<th>Fraction</th>
<th>UV (nm)</th>
<th>[M-H]</th>
<th>[M+H]^+</th>
<th>Proposed compound</th>
<th>Quantity (mg/g of crude propolis)</th>
</tr>
</thead>
<tbody>
<tr>
<td>11</td>
<td>19.18</td>
<td>MeOH</td>
<td>300, 330</td>
<td>353</td>
<td>355.1</td>
<td>3-O-Caffeoylquinic acid</td>
<td>0.16 0.38 0.11 0.17 0.05 0.25</td>
</tr>
<tr>
<td>12</td>
<td>21.65</td>
<td>MeOH</td>
<td>300, 330</td>
<td>352.9</td>
<td>352.2</td>
<td>5-O-Caffeoylquinic acid</td>
<td>1.96 4.14 1.83 2.93 0.60 3.18</td>
</tr>
<tr>
<td>13</td>
<td>25.82</td>
<td>MeOH</td>
<td>312</td>
<td>nd</td>
<td>355.1</td>
<td>4-O-Caffeoylquinic acid</td>
<td>0.61 0.76 0.45 0.71 0.08 0.35</td>
</tr>
<tr>
<td>14</td>
<td>37.15</td>
<td>MeOH</td>
<td>290, 325</td>
<td>519.1</td>
<td>521.3</td>
<td>didihydrocaffeoylquinic acid</td>
<td>0.67 0.92 1.50 0.21 0.32</td>
</tr>
<tr>
<td>15</td>
<td>39.62</td>
<td>MeOH</td>
<td>300, 330</td>
<td>515.1</td>
<td>nd</td>
<td>3,5-di-O-Caffeoylquinic acid</td>
<td>7.41 11.96 4.78 11.38 1.98 8.24</td>
</tr>
<tr>
<td>16</td>
<td>34.07</td>
<td>EtOAc</td>
<td>310</td>
<td>162.9</td>
<td>nd</td>
<td>p-Coumaric acid</td>
<td>7.43 6.20 7.67 2.89 1.41 1.96</td>
</tr>
<tr>
<td>17</td>
<td>50.09</td>
<td>MeOH</td>
<td>300, 330</td>
<td>515.1</td>
<td>517.2</td>
<td>4,5-di-O-Caffeoylquinic acid</td>
<td>10.52 19.35 6.97 16.78 2.89 13.79</td>
</tr>
<tr>
<td>18</td>
<td>53.14</td>
<td>MeOH</td>
<td>295, 325</td>
<td>529.2</td>
<td>531.3</td>
<td>3-O-Feruloyl-5-O-Caffeoylquinic acid</td>
<td>0.56 0.35 0.38 0.96 0.02 0.45</td>
</tr>
<tr>
<td>19</td>
<td>61.90</td>
<td>MeOH</td>
<td>300, 330</td>
<td>677.1</td>
<td>679.3</td>
<td>3,4,5-tri-O-Caffeoylquinic acid</td>
<td>3.13 3.61 1.99 4.46 1.07 3.00</td>
</tr>
<tr>
<td>20</td>
<td>65.90</td>
<td>EtOAc</td>
<td>290</td>
<td>301.1</td>
<td>303.1</td>
<td>Methoxypinobanksin</td>
<td>4.22 3.96 2.52 1.55 0.04 0.41</td>
</tr>
<tr>
<td>21</td>
<td>71.08</td>
<td>EtOAc</td>
<td>268, 365</td>
<td>315</td>
<td>317.1</td>
<td>Isorhamnetin</td>
<td>6.13 4.76 2.25 2.40 0.58 0.78</td>
</tr>
<tr>
<td>22</td>
<td>74.35</td>
<td>EtOAc</td>
<td>265, 350</td>
<td>299</td>
<td>301.2</td>
<td>Luteolin-5-methyl ether</td>
<td>0 0 0 0 2.95 2.97</td>
</tr>
<tr>
<td>5</td>
<td>78.43</td>
<td>EtOAc</td>
<td>315</td>
<td>231</td>
<td>nd</td>
<td>4-Hydroxy-3-prenylcinnamic acid (drupanin)</td>
<td>0.73 1.15 0.58 0.17 0.04 0.32</td>
</tr>
<tr>
<td>10</td>
<td>81.54</td>
<td>EtOAc</td>
<td>275sh. 318</td>
<td>315</td>
<td>nd</td>
<td>3-Prenyl-4-(2-methylpropionyl-oxy)-cinnamic acid</td>
<td>0.17 0.29 0.06 0.02 0 0</td>
</tr>
<tr>
<td>23</td>
<td>82.88</td>
<td>EtOAc</td>
<td>268, 365</td>
<td>299</td>
<td>301.2</td>
<td>Kaempferol</td>
<td>15.31 10.18 11.57 5.59 1.92 3.07</td>
</tr>
<tr>
<td>8</td>
<td>87.08</td>
<td>EtOAc</td>
<td>320</td>
<td>315.1</td>
<td>nd</td>
<td>3-hydroxy-2,2-dimethyl-8-prenylchromane-6-propenoic acid</td>
<td>0.13 0.19 0.16 0.05 0.09 0.10</td>
</tr>
<tr>
<td>9</td>
<td>91.49</td>
<td>EtOAc</td>
<td>315</td>
<td>299.1</td>
<td>nd</td>
<td>4-Hydroxy-3,5-diprenylcinnamic acid (artepillin C)</td>
<td>1.02 0.86 0.35 0.08 0.15 0.30</td>
</tr>
<tr>
<td>24</td>
<td>97.18</td>
<td>EtOAc</td>
<td>290, 330</td>
<td>363.1</td>
<td>nd</td>
<td>3-Prenyl-4-(dihydrocinnamoyl-oxy)-cinnamic acid (baccharin)</td>
<td>0.10 0 0.11 0 0.09</td>
</tr>
</tbody>
</table>

Table 3. Parameters of standard curves used for quantification of major compounds of samples from Brazilian propolis.

<table>
<thead>
<tr>
<th>Standard</th>
<th>Range (µg)</th>
<th>equation</th>
<th>R²</th>
</tr>
</thead>
<tbody>
<tr>
<td>Quercetin</td>
<td>0.1 - 2</td>
<td>y=0.0005x</td>
<td>0.9958</td>
</tr>
<tr>
<td>Chlorogenic acid</td>
<td>0.05 - 6.0</td>
<td>y=0.0002x</td>
<td>0.9982</td>
</tr>
<tr>
<td>p-Coumaric acid</td>
<td>0.01 - 1.5</td>
<td>y=0.00006x</td>
<td>0.9939</td>
</tr>
</tbody>
</table>

Other prenylated phenylpropanoids occurring as minor constituents of the samples analyzed are 7 and 10. Among the caffeoylquinic acids, 17 (a dicaffeoylquinic acid) predominates. In addition to caffeoylquinic acids, flavonoids, such as methoxypinobanksin (20), isorhamnetin (21) and kaempferide (23) are also relevant constituents, mainly with respect to the samples from Minas Gerais (A-D). The simple phenylpropanoid p-coumaric acid (16) was also an important constituent of all samples from Minas Gerais (Table 2).

Flavonoids have been regarded as minor constituents of Brazilian propolis. The data given in Table 2, however, indicate that another plant is the origin of this flavonoid. The equations and coefficients of the standard curves for the HPLC quantitative analyses are shown in Table 3. The contents of the constituents of the samples analyzed are shown in Table 2.

Relatively low contents of the phenylpropanoids drupanin (5), artepillin C (9) and baccharin (24) characterize the samples analyzed.
are major constituents of Brazilian green propolis,\(^2,3\) the quantitative analysis of the present study indicates that dicaffeoylquinic acids are the most abundant compounds of this type of propolis.

**CONCLUSIONS**

Although both marker compounds of Brazilian green propolis were detected in the samples from Minas Gerais and Paraná, the qualitative and quantitative analyses indicated that the samples from the two localities are chemically quite distinct and that quantitative aspects should be taken into account to address the complex problem of propolis standardization.

Plants of aecerim-do-campo are not abundant in Paraná, and thus other sources (such as poplar and an unknown source of 22) probably complement the provision of resin for propolis production in this state. Therefore, samples of propolis from Paraná likely have a more complex botanical origin than the samples from Minas Gerais.

Quantitative analysis might contribute toward a revision of the traditional concept that prenylated phenylpropanoids are the most abundant constituents of Brazilian green propolis.

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**REFERENCES**